Ultrasonically Induced Conjugate Addition of Iodides to Electron-Deficient Olefins and Its Application to the Synthesis of Side-Chain Analogs of the Hormone 1α , 25-Dihydroxyvitamin D₃¹

José Pérez Sestelo, José L. Mascareñas, Luis Castedo, and Antonio Mouriño*

Departamento de Química Orgánica y Sección de Alcaloides del CSIC, Universidad de Santiago de Compostela, 15706 Santiago de Compostela, Spain

Received July 13, 1992

A versatile method for the rapid construction of fragments related to the upper part of vitamin D_3 from the Lythgoe-Inhoffen diol is described. The key feature of the strategy is a new zinc-copperinduced conjugate addition of iodo triflate 10a to electron-deficient olefins under sonochemical aqueous conditions. These fragments are rapidly and efficiently transformed via the dienyne convergent approach to several derivatives of the hormone 1α , 25-(OH)₂-D₃ modified at C-25.

Introduction

Vitamin D_3 (1a) is a prohormone classically associated with homeostasis of calcium and phosphorus through the regulation of intestinal calcium absorption and bone calcium mobilization.² It is now well-established that these biological effects of vitamin D₃ occur as a consequence of its sequential metabolism to 25-hydroxyvitamin D_3 (1b) in the liver and to the hormone 1α ,25-dihydroxyvitamin D_3 (1c) in the kidney. Like other steroid hormones, the mode of action of 1c on target cells is mediated by specific receptors.^{2,3}



Investigations carried out during the last decade have shown that 1α , 25-dihydroxyvitamin D₃ is involved in the regulation of the function of a broader range of cells than those of bone and intestine.⁴ It has been found that the hormone induces the terminal differentiation and supresses the proliferation of human myeloid leukemic cells,⁵ epidermal keratinocytes,⁶ and several types of cancer cell⁷ and alters the immune response of lymphocytes.⁸ These findings have enormously stimulated research aimed at evaluating the clinical possibilities of the hormone and its derivatives in the treatment of cancer, psoriasis, and immune disorders.⁵⁻⁹ The cellular mechanisms by which these effects are brought about are not yet fully understood, but is believed that both classical nuclear receptormediated regulation of gene transcription and nongenomic pathways may be involved.¹⁰

A key issue with regard to the spectrum of biological effects of 1α , 25-(OH)₂-D₃ (1c) is whether it is possible to design analogs which are capable of inducing selective or differential biological responses, e.g., promotion of cell differentiation without stimulation of calcium activity.¹¹ A growing amount of research is being directed to this end, and the results reported so far indicate that sidechain modifications are especially effective for modulating the selectivity and activity of the hormone.^{10,12} It is

⁽¹⁾ Part of this work was presented at the Second European Symposium on Sonochemistry, Gargnano, Italy, Sept 1991. (2) (a) Norman, A. W. Vitamin D, the Calcium Homeostatic Steroid

⁽a) (a) Ivolman, A. w. recomme D, the Calcium homeostatic Sterold Hormone; Academic Press: New York, 1979. (b) Norman, A. W.; Roth, J.; Orci, L. Endocrine Rev. 1982, 3, 311. (c) DeLuca, H. F.; Schnoes, H. K. Ann. Rev. Biochem. 1983, 52, 411. (d) DeLuca, H. F.; Schnoes, H. K. Ann. Rep. Med. Chem. 1984, 19, 179. (e) Jones, G. Steroids 1987, 49, 1. (3) Haussler, M. R. Annu. Rev. Nutr. 1986, 6, 527.
(4) (a) Holick M. F. Kidm. Int. 1987, 29, 219. (b) Optimum V. K. Deluca, (d) (a) Holick M. F. Kidm. 1981, 1992,

 ^{(4) (}a) Holick, M. F. Kida. Int. 1987, 32, 912. (b) Ostrem, V. K.; DeLuca,
 H. F. Steroids 1987, 49, 73. (c) DeLuca, H. F. FASEB J. 1988, 2, 224. (d) Minghetti, P. P.; Norman, A. W. FASEB J. 1988, 2, 3043. (e) Reichel, H.; Koeffler, H. P.; Norman, A. W. N. Engl. J. Med. 1989, 320, 980. (f) Manolagas, S. C.; Hustmyer, F. G.; Yu, X. P. Kidney Int. 1990, 38 (Supp. 29), S9-S16. (g) Carson-Jurica, M. A.; Schrader, W. T.; O'Malley, B. W. Endocrine Rev. 1990, 11, 201. (h) DeLuca, H. F.; Burmester, J.; Darwish,

<sup>Endocrine Rev. 1990, 11, 201. (h) DeLuca, H. F.; Burmester, J.; Darwish, H.; Krisinger, J. Comprehensive Medicinal Chemistry; Pergamon Press: New York, 1990; Vol. 3, pp 1129.
(5) (a) Abe, E.; Miyaura, C.; Sakagami, H.; Takeda, M.; Konno, K.; Yamazaki, T.; Yoshiki, S.; Suda, T. Proc. Nat. Acad. Sci. U.S.A. 1981, 78, 4990. (b) Honna, Y.; Hozumi, M.; Abe, E.; Konno, K.; Fukushima, M.; Hata, S.; Nishii, Y.; DeLuca, H. F.; Suda, T. Proc. Nat. Acad. Sci. U.S.A. 1983, 80, 201. (c) MacLaughlin, J. A.; Gange, W.; Taylor, D.; Smith, E.; Holick, M. F. Proc. Nat. Acad. Sci. U.S.A. 1985, 82, 5409. (d) Suda, T. Proc. Soc. Exp. Biol. Med. 1989, 191, 215. (e) Abe, J.; Takito, T. N.; Miyaura, C.; Suda, T.; Nishii, Y. Enderinology 1989. 124, 2645.</sup> T. N.; Miyaura, C.; Suda, T.; Nishii, Y. Endocrinology 1989, 124, 2645.

^{(6) (}a) Kurok, T. J. Invest. Dermatol. 1985, 84, 459. (b) Smith, E. L.; Walworth, N. C.; Holick, M. F. J. Invest. Dermatol. 1986, 86, 709.

^{(7) (}a) Eisman, J. A.; Barkla, D. H.; Tutton, P. J. M. Cancer Res. 1987 47, 21. (b) Eisman, J. A.; Koga, M.; Sutherland, R. L.; Barkla, D. H.; Tutton, P. J. M. Reference 5d, pp 221-226.

^{(8) (}a) Tsoukas, C. D.; Provvedini, D. M.; Manolagas, S. C. Science (Washington, D.C.) 1984, 224, 1438. (b) Rigby, W.F.C. Immunol. Today 1988, 9, 54. (c) Manolagas, S. C.; Hustmyer, F. G.; Yu, X. P. ref 5d, pp 238–245. (d) Yu, X. P.; Hustmyer, F. G.; Garvey, W. T.; Manolagas, S. C. Proc. Nat. Acad. Sci. U.S.A. 1991, 88, 8347.

⁽⁹⁾ The hormone and its derivatives are remarkably effective in the treatment of psoriasis: (a) Kragballe, K. Arch. Dermatol. 1989, 125, 1647. (b) Holick, M. F. Proc. Soc. Exp. Biol. Med. 1989, 191, 246-257. (c) Thavajarah, M.; Evans, D. B.; Binderup, L.; Kanis, J. A. Biochem. Biophys. Res. Commun. 1990, 171, 1056. (10) Norman, A. W.; Bouillon, R.; Thomasset, M., Eds.; Vitamin D:

Gene Regulation, Structure Function Analysis and Clinical Application; Walter de Gruyter and Co: Berlin, 1991.

⁽¹¹⁾ Therapeutically effective doses of the hormone induce

⁽¹¹⁾ Therapeutically effective doses of the hormone induce hypercalcemia: Koeffler, H. P.; Hirji, K.; Itri, L. The Southern California Leukemia Group; Cancer Treat. Rep. 1985, 69, 1399.
(12) (a) Ikekawa, N. Med. Chem. Rev. 1987, 7, 333. (b) Abe, J. Morikawa, M.; Miyamoto, K.; Kaiho, S.; Fukushima, M.; Miyaura, C.; Abe, E.; Suda, T.; Nishii, Y. FEBS Lett. 1987, 226, 58. (c) Ostrem, V. K.; Lau, W. F.; Lee, S. H.; Perlman, K.; Prahl, J.; Schnoes, H. K.; DeLuca, L. H. F. J. Biol. Chem. 1987, 262, 14154. (d) Shinay, S.; Dentrider, L. L. H. F. J. Biol. Chem. 1987, 262, 14164. (d) Shiuey, S. J.; Partridge, J. J.; Uskokovic, M. R. J. Org. Chem. 1988, 53, 1040. (e) Zhou, J.; Norman, A. W.; Lubbert, M.; Collins, E. D.; Uskokovic, M. R.; Koeffler, H. P. Blood 1989, 74, 82. (f) DeLuca, H. F.; Ikekawa, N.; Tanaka, D. US Pat. 1989, 4, 851, 400. (g) DeLuca, H. F.; Schnoes, H. K.; Perlman, K. L.; Kutner, A. US Pat. 1989, 4, 851, 401. (h) Perlman, K. L.; Sicinski, R. R.; Schnoes,
 H. K.; DeLuca, H. F. Tetrahedron Lett. 1990, 31, 1823. (i) Figadère, B.;
 Norman, A. W.; Henry, H. L.; Koeffler, H. P.; Zhou, J.-Y.; Okamura, W.
 H. J. Med. Chem. 1991, 34, 2452. (i) Kissmeyer, A.-M.; Binderup, L. W.; Biochem. Pharmacology 1991, 41, 1601. (k) Zhou, J.-Y.; Norman, A Akashi, M.; Chen, D.-L.; Uskokovic, M. R.; Aurrecoechea, J. M.; Dauben, W. G.; Okamura, W. H.; Koeffler, H. P. Blood 1991, 78, 75. (1) Calverley, M. J. Tetrahedron 1987, 43, 4609.



therefore of interest to develop flexible methods allowing the synthesis of a wide variety of side-chain analogs that could be screened for clinical application and/or used as biochemical tools for investigating the molecular mechanisms underlying the biological actions.

We have recently reported a short, flexible route to 25,26,27-trisnor-24-(methoxycarbonyl)vitamin D₃. The key steps were a sonochemical-induced conjugate addition of iodide 6b to methyl acrylate and a Wittig-Horner coupling between the keto ester 7 and a known vitamin D ring-A phosphine oxide synthon.¹³ Here, we report the application of the sonochemical methodology in conjunction with the dienyne approach to the vitamin D triene¹⁴ for the preparation of the more important 1α -hydroxylated derivatives. Our efforts have led to the development of a versatile route for the synthesis of the hormone 1c and several analogs modified at C-25 of potential biomedical interest (14-18). The key feature of the strategy is a new zinc-copper-induced conjugate addition of the iodo triflate 10a to methyl acrylate and other α,β -unsaturated compounds under sonochemical aqueous conditions.

Results and Discussion

The synthetic plan for the preparation of the 1α -hydroxy derivatives shown above is based on our recently improved convergent dienyne strategy (Scheme I).¹⁴ The availability of ring-A enyne 5¹⁵ makes this route especially convenient.

Synthesis of Ester 13. Although the required triflate 8 can be prepared from the known keto ester 7,13 the best conditions found to carry out this transformation [slow addition of 7 to a cold $(-90 \,^{\circ}\text{C})$ solution of LDA $(1.1 \, \text{equiv})$ in THF and subsequent quenching with excess N-phenyltriflimide] gave 8 in only 51% yield (Scheme II).

We then envisaged the possibility of forming the vinyl triflate before the introduction of the side chain so as to avoid problems arising from the presence of acidic hydrogens in the latter. It was reasoned that the mild conditions used for sonochemical conjugate addition or alkyl halides to electron-deficient olefins might permit the construction of the side chain in the presence of the vinyl triflate.^{13,18} Iodide 6b, previously synthesized from



the Inhoffen-Lythgoe diol 6a by a two-step procedure,^{14c} was better prepared in 95% yield by reaction of **6a** with Ph₃P and I₂ at 0 °C following Samuelson's procedure.¹⁶ Iodo alcohol 6b was efficiently oxidized with PDC to iodo ketone 9 (Scheme III), and the kinetic enolate prepared by treatment of this ketone with LDA at low temperature was trapped with N-phenyltriflimide to give triflate 10a (86% for two steps). Interestingly, sonication of a mixture of 10a, Zn, CuI, and methyl acrylate in 7:3 EtOH/H₂O at room temperature afforded ester 8 in a satisfactory 65% yield.^{13,17} Most of the remaining material separated was identified as the protonated product 10b. Although the mechanism of the reaction is still not clear, studies by Luche and co-workers indicate that the reaction follows most probably a radical pathway, the key step being a sonochemically promoted single-electron transfer from the metal to the C-X bond.¹⁸ The compatibility of the sonochemical reaction with the vinyl triflate, which would not be left intact by most current organometallic methods for C-C bond formation, expands the scope and usefulness of this procedure and further illustrates the high potential of ultrasound in synthesis.¹⁹

Palladium-catalyzed coupling between vinyl triflate 8 and enyne 5 furnished dienyne 11 in 86% yield. Partial hydrogenation of the dienyne in the presence of Lindlar catalyst poisoned with quinoline, with careful monitoring of the reaction to avoid overreduction of the triple bond. afforded 12 in 95% yield. The previtamin 12 was refluxed in isooctane for 4 h to bring about equilibration to the vitamin 13.20

Synthesis of C-25 Dialkylated Derivatives of the Hormone 1c. With ester 13 at hand, the preparation of 1c, its labeled counterpart 14, and C-25 dialkylated derivatives was straightforward. As shown in Table I the hormone 1c as well as the hexadeuteriated derivative 14 were efficiently obtained by treatment of ester 13 with the corresponding Grignard reagent and subsequent removal of the silyl groups with TBAF. The number of steps and overall yield of the synthesis of 1c (eight steps from the

⁽¹³⁾ Mascareñas, J. L.; Pérez-Sestelo, J.; Castedo, L.; Mouriño, A. Tetrahedron Lett. 1991, 32, 2813.

^{(14) (}a) Castedo, L.; Mouriño, A.; Sarandeses, L. A. Tetrahedron Lett. 1986, 27, 1523. (b) Castedo, L.; Mascareñas, J. L.; Mouriño, A.; Sarandeses, . A. Tetrahedron Lett. 1988, 29, 1203. (c) Mascareñas, J. L.; Sarandeses, L. A.; Castedo, L.; Mouriño, A. Tetrahedron 1991, 47, 3485.

 ^{(15) (}a) Castedo, L.; Mascarefias, J. L.; Mouriño, A. Tetrahedron Lett.
 1987, 28, 2099. (b) Okamura, W. H.; Aurrecoechea, J. M.; Gibbs, R. A.; Norman, A. W. J. Org. Chem. 1989, 54, 4072. (c) Hatakeyama, S.; Numata, H.; Osanai, K.; Takano, S. J. Org. Chem. 1989, 54, 3515.

⁽¹⁶⁾ Garegg, P. J.; Samuelsson, B. J. Chem. Soc., Chem. Commun. 1979, 978.

⁽¹⁷⁾ A control experiment without sonication indicated no progress of

<sup>the reaction after stirring for 26 h.
(18) (a) Petrier, C.; Dupuy, C.; Luche, J. L. Tetrahedron Lett. 1986, 27, 3149.
(b) Luche, J. L.; Allavena, C. Tetrahedron Lett. 1988, 29, 5369.</sup> (c) Luche, J. L.; Allavena, C.; Petrier, C.; Dupuy, C. Tetrahedron Lett.
 1988, 29, 5373. (d) Dupuy, C.; Petrier, C.; Sarandeses, L.; Luche, J. L. Synth. Commun. 1991, 21, 643. (19) (a) Einhorn, C.; Einhorn, J.; Luche, J. L. Synthesis 1989, 787. (b)

Ley, S. V.; Low, C. M. R. In Ultrasound in Synthesis; Springer Verlag: 1989.

⁽²⁰⁾ Napoli, J. L.; Mellon, W. S.; Fivizzani, M. A.; Schnoes, H. K.; DeLuca, H. F. Biochemistry 1980, 19, 2515.



Table I. Dialkylation of Ester 13

entry	organomet reagent ^a	product ^b	% yield ^c
1	IMgCH ₃	1c	73
2	IMgCD ₃	14	72
3	cyclopropyllithium	15	65
4	tert-butyllithium	16	68
5	phenyllithium	17	71

^a 2.2 equiv. ^b These are the products obtained after removal of the silyl groups with TBAF. ^c Overall yield for two steps.

Inhofen–Lythgoe diol, 41% overall yield) compare favorably with previously reported approaches.^{14c,21}

The remarkably low calcemic and potent antiproliferative properties exhibited by several 26,27-dialkylated derivatives of the hormone^{10,22} prompted us to use ester 13 to prepare derivatives of this kind. The methods previously described for the synthesis of these 26,27dialkylated derivatives generally involve long linear approaches.²² Analogs 15, 16, and 17, in which the 26,27-CH₃ groups of the hormone are replaced by cyclopropyl, *tert*-butyl, and phenyl groups, respectively, were readily synthesized by reaction of 13 with the corresponding lithium carbanions and subsequent desilylation (Table I). The biological activity of these compounds is now being tested, and the results will be reported elsewhere. We hope that the data obtained will confirm the therapeutical possibilities of this kind of compounds and will contribute to understanding the relation between the spatial environment of the 25-OH group and biological activity.

Sonochemical Addition of Iodo Triflate 10a to other Olefins. With the purpose of applying our approach to

Table II. Sonochemical Conjugate Additions

10a	Zn, Cul EtOH/H ₂ O	тю	- 2
Z^a	adduct	time (min)	yield ^b (%)
CO ₂ CH ₃	8	45	65
COCH ₃	19	35	73
CN	20	40	65
$CONH_2$	21	75	35
SO ₂ CH ₃	22	15	72
SOPh	23	70	45

^a 5-20 equiv in excess of the olefin are used. ^b Isolated yield.

the synthesis of a wider range of side-chain analogs of the hormone and to assess the scope and potential of the sonochemical reaction described above, the addition of iodide 10a to other types of olefins was studied. The results are summarized in Table II. Reaction of 10a with methyl vinyl ketone, acrylonitrile, and acrylamide gave the adducts in synthetically useful yields. Reaction with olefins having sulfone or sulfoxide as electron-withdrawing groups was also satisfactory. The efficiency of the addition to the vinyl sulfoxide is particularly noteworthy since other carbon nucleophiles are usually unreactive toward this unsaturated functionality.²³

Synthesis of Sulfonyl Derivative 18. It has been reported that introduction of heteroatoms into the side chain of hormone 1c can afford biologically interesting analogs.^{12b,24} However, few derivatives with heteroatoms in place of carbon atoms at C-25 have hitherto been synthesized.²⁵ Having shown that the above sonochemical reaction of the key iodo triflate 10a allows rapid and practical construction of side chains with heteroatoms at that position, we validated the dienyne route to the corresponding vitamin D analogs by using it to prepare

^{(21) (}a) Paaren, H. E.; DeLuca, H. F.; Schnoes, H. K. J. Org. Chem.
1980, 45, 3253. (b) Pardo, R.; Santelli, M. Bull. Soc. Chim. Fr. 1985, 98.
(c) Baggiolini, E. G.; Iacobelli, J. A.; Hennessy, B. M.; Batcho, A. D.;
Sereno, J. F.; Uskokovic, M. R. J. Org. Chem. 1986, 51, 3098. (d) Andrews,
D. R.; Barton, D. H. R.; Hesse, R. H.; Pechet, M. M. J. Org. Chem. 1986, 51, 4819. (e) Wilson, S. R.; Venkatesan, A. M.; Augelli-Szafran, C. E.;
Yasmin, A. Tetrahedron Lett. 1991, 32, 2339.

<sup>Yasmin, A. Tetrahedron Lett. 1991, 32, 2339.
(22) (a) Sai, H.; Takatsuko, S.; Hara, N.; Ikekawa, N. Chem. Pharm.
Bull. 1985, 33, 878. (b) Ostrem, V. K.; Tanaka, Y.; Prahl, J.; DeLuca, H.
F.; Ikekawa, N. Proc. Nat. Acad. Sci. U.S.A. 1987, 84, 2610. (c) Ikekawa,
N.; Eguchi, T.; Hara, N.; Takatsuto, S.; Honda, A. Mori, Y.; Otomo, S.
Chem. Pharm. Bull. 1987, 35, 4362. (d) Eguchi, T.; Takatsuko, S.; Hara,
N.; Ikekawa, N. Chem. Pharm. Bull. 1988, 36, 2303. (e) Gull, H. S.;
Londowski, J. J.; Corradino, R. A.; Zinsmeister, A.; Kumar, R. J. Med.
Chem. 1990, 33, 480. (f) Eguchi, T.; Ikekawa, N.; Sumitani, K.; Kumegawa,
M.; Higuchi, S.; Otomo, S. Chem. Pharm. Bull. 1980, 38, 1246.</sup>

⁽²³⁾ Iwata, C.; Maezaki, N.; Kurumada, T.; Fukuyama, H.; Sugiyama, K.; Imanishi, T. J. Chem. Soc., Chem. Commun. 1991, 1408.

⁽²⁴⁾ Neef, G.; Steinmeyer, A. Tetrahedron Lett. 1991, 32, 5073.

^{(25) (}a) Onisko, B. L.; Schnoes, H. K.; DeLuca, H. F. Bioorg. Chem.
1980, 9, 187. (b) Dauben, W. G.; Ollmann, Jr. R. R.; Funhoff, A. S.;
Leung, S. S.; Norman, A. W.; Bishop, J. E. Tetrahedron Lett. 1991, 32, 4643.



sulfone 18. It is expected that the presence of the polar sulfone group will alter the transport and binding properties of the compound and that this may affect its biological behavior.

Palladium-catalyzed coupling of triflate 22 with enyne 5 gave dienyne 24 in a moderate 65% yield (Scheme IV). Partial hydrogenation, thermal isomerization, and removal of the silyl groups afforded the hormone analog 18 (78% yield for three steps). The application of our approach to the synthesis of other analogs of the hormone and the biological evaluation (antiproliferative and calcemic activity as well as receptor affinities) of all these compounds will be reported elsewhere.

Conclusion

A short, versatile approach to side-chain derivatives of the hormone $1\alpha,25$ -(OH)₂-D₃ modified at C-25 has been described. The key feature of the strategy is the ultrasonically-induced conjugate addition of iodo trilfate 10a to electron-deficient olefins under aqueous conditions. The mildness and simplicity of this method and the fact that, unlike current methods for carbon-carbon bond formation it is compatible with the vinyl triflate group, illustrates the potential of sonochemical procedures in organic synthesis.

In contrast to lengthy linear approaches previously reported, the route described here is especially efficient for preparation of C-25 dialkylated and C-25 heterosubstituted derivatives of hormone 1c. It is our expectation that the information obtained from assays of the biological activity and binding properties of such analogs will contribute to our understanding of the mode of action of the hormone and will make feasible the design of new, more effective analogs.

Experimental Section

General. All dry solvents were freshly distilled under argon from the appropriate drying agent before use. Et₂O, benzene, toluene, and THF were distilled from sodium/benzophenone. CH₂Cl₂ was distilled from P₂O₅, and hexanes from LAH. Absolute MeOH was distilled from Mg turnings. Py was distilled from KOH. *i*-Pr₂NH, Et₃N, and isooctane were distilled and then redistilled from CaH₂. DMF was distilled from P₂O₅ under vacuum and stored under type 4A molecular sieves. All reactions were conducted in dry solvents under argon atmosphere unless otherwise stated. Kugelrohr distillation boiling points refer to the external air bath temperature (at). Boiling points and melting points (open capillary tubes) are uncorrected. CuI was purified by the method of Kauffman²⁶ and Zn as recommended in the book by Fieser and Fieser.²⁷ Flash chromatography was performed by Still's method.²⁸ TLC was performed on silica gel plates, and components were located by observation under UV light and/or by treating the plates with a phosphomolybdic acid reagent followed by heating. Sonications were carried out in a Bandelin 120–240 W 35-kHz cleaning bath (Ultrasonic). Dryings were carried out with anhydrous Na₂SO₄. Concentrations were carried out in a rotary evaporator.

Full spectral and analytical data and the ¹H NMR and ¹³C NMR spectra are given in the supplementary material. Essential ¹H NMR and ¹³C NMR data (including carbon types determined from DEPT experiments) are also presented in the Experimental Section. Satisfactory combustion analysis and/or high-resolution mass spectra were obtained for all new compounds.

Des-A,B-22-iodo-23,24-dinorcholan-86-ol (6b). Ph₃P (6.5 g, 27.76 mmol) and imidazole (4.8 g, 70.7 mmol) were added to a solution of Inhoffen–Lythgoe diol 6a²⁹ (5 g, 23.6 mmol) in THF (100 mL). The suspension was cooled to -20 °C, and I₂ (6.28 g, 24.76 mmol) was added. After being stirred for 15 min, the reaction mixture was warmed to rt, further stirred for 1.5 h, cooled to 0 °C, and poured into saturated aqueous NaHCO₃ (50 mL). The mixture was extracted with $Et_2O(2 \times 60 \text{ mL})$, and the extract was washed with saturated aqueous $Na_2S_2O_3$ and H_2O , dried, and filtered. Concentration afforded a residue which was purified by flash chromatography (8% EtOAc/hexanes) to give 7.32 g of 6b (96%).14c The white solid was crystallized from EtOAc/ hexanes (mp 51 °C): ¹³C NMR δ 69.1 (C-8), 55.9, 52.3, 41.8 (C), 40.0 (CH2), 36.3, 33.5 (CH2), 26.4 (CH2), 22.3 (CH2), 21.1 (CH2), 20.6, 17.3 (CH₂), 14.3. Anal. Calcd for C₁₃H₂₃OI: C, 48.46; H, 7.19. Found: C, 48.58; H, 7.36.

Des-*A*,*B***-22-iodo-23,24-dinorcholan-8-one (9).** PDC (8.66 g, 24 mmol) was added to a solution of the alcohol **6b** (3.99 g, 12 mmol) in CH₂Cl₂ (50 mL). The mixture was stirred for 6 h at rt. Et₂O (60 mL) was added, and the resulting suspension was stirred for 15 min and filtered through a short pad of Celite and silica gel. The filtrate was washed with brine (35 mL), dried, filtered, and concentrated. The residue was purified by flash chromatography (10% EtOAc/hexanes) to give iodo ketone 9 (3.57 g, 90%). The white solid was crystallized from Et₂O/hexanes (mp 65 °C): ¹H NMR δ 3.31 (1 H, dd, $J \sim 9.7$, 2.5 Hz, H-22), 3.19 (1 H, dd, $J \sim 9.7$, 4.8 Hz, H-22), 1.06 (3 H, d, $J \sim 5.8$ Hz, C₂₁-CH₃), 0.68 (3 H, s, C₁₈-CH₃); ¹³C NMR δ 211.1 (CO), 61.3, 55.6, 49.3 (C), 40.6 (CH₂), 38.4 (CH₂), 36.1, 26.6 (CH₂), 23.7 (CH₂), 20.6, 20.3 (CH₂), 18.7 (CH₂), 13.0. Anal. Calcd for C₁₃H₂₁OI: C, 48.76; H, 6.61. Found: C, 48.68; H, 6.47.

Procedure for the Preparation of Vinyl Triflates: Des-A,B-22-iodo-23,24-dinorchol-8-enyl Trifluoromethanesulfonate (10a). Lithium diisopropylamide (LDA) was prepared by addition of i-Pr₂NH (0.54 mL, 3.9 mmol) to a cooled (-78 °C) solution of n-BuLi in hexanes (1.43 mL, 2.50 M, 3.5 mmol). After the mixture was stirred for 10 min, a white precipitate appeared. The mixture was diluted with THF (4 mL), stirred at 0 °C for 30 min, and cooled to -78 °C. A solution of ketone 9 (1 g, 3.12 mmol) in THF (14 mL) was slowly added (35 min), followed by a solution of N-phenyltriflimide (1.225 g, 3.43 mmol) in THF (4 mL). The mixture was stirred for 2 h at -78 °C. After being warmed to 0 °C, the reaction was quenched by addition of a few drops of MeOH and water. Concentration gave a crude product which was diluted with EtOAc/hexanes (30 mL, 1:3), washed with brine (20 mL), dried, filtered, and concentrated. The resulting viscous residue was purified by flash chromatography (2% EtOAc/hexanes) affording 1.29 g of iodo triflate 10a (92%, colorless oil): ¹H NMR δ 5.59 (1 H, dd, $J \sim$ 6.9, 3.5 Hz, H-9), 3.32 (1 H, dd, J ~ 9.7, 2.6 Hz, H-22), 3.18 (1 H, dd, J ~ 9.7, 5.1 Hz, H-22), 2.52 (1 H, m, H-14), 2.30 (2 H, m, two H-11), 1.06 (3 H, d, J ~ 5.8 Hz, C₂₁-CH₃), 0.8 (3 H, s, C₁₈-CH₃); ¹³C NMR δ 149.4

⁽²⁶⁾ Kauffman, G. B.; Teter, L. A. Inorg. Synth. 1963, 7, 9.

⁽²⁷⁾ Fieser, L. F.; Fieser, M. Reagents for Organic Synthesis; John Wiley & Sons: New York, 1967; Vol. 1, p 1292.

⁽²⁸⁾ Still, W. C.; Kahn, M.; Mitra, A. J. Org. Chem. 1978, 43, 2923.
(29) (a) Lythgoe, B.; Roberts, D. A.; Waterhose, I. J. Chem. Soc., Perkin Trans. 1 1977, 2608. (b) Leyes, G. A.; Okamura, W. H. J. Am. Chem. Soc.
1982, 104, 6099. (c) Sardina, F. J.; Mouriño, A.; Castedo, L. J. Org. Chem.
1986, 51, 1264. (d) Barrack, S. A.; Gibbs, R. A.; Okamura, W. H. J. Org. Chem. 1988, 53, 1790. (e) Mascareñas, J. L.; Sarandeses, L. A.; Castedo, L.; Mouriño, A. Tetrahedron 1991, 47, 3485. (f) Craig, A. S.; Norman, A. W.; Okamura, W. H. J. Org. Chem. 1992, 57, 4374.

(C-8), 121.0 (CF₃), 116.0 (C-9), 53.4, 49.7, 45.0 (C), 36.7, 34.4 (CH₂), 27.4 (CH₂), 23.6 (CH₂), 21.2 (CH₂), 20.6, 19.8 (CH₂), 11.9. Anal. Calcd for $C_{14}H_{20}O_3F_3SI$: C, 37.18; H, 4.46. Found: C, 37.43; H, 4.36.

Procedure for Sonochemical Conjugate Additions: Methyl Des-A, B-8-[(trifluoromethanesulfonyl)oxy]chol-8ene-24-carboxylate (8). A suspension of CuI(201 mg, 1.06 mmol) and Zn (161 mg, 2.5 mmol) in EtOH/H₂O (6 mL, 7:3, previously deoxygenated under reduced pressure) was sonicated for 5 min. The olefin, in this case methyl acrylate (637 μ L, 7.8 mmol, freshly distilled under vacuum), and a solution of iodide 10a (160 mg, 0.35 mmol) in EtOH/H₂O (1 mL, 7:3) were successively added, and the resulting mixture was sonicated for 40 min. Dilution with Et₂O (15 mL) and filtration through a short pad of celite (washed repeatedly with Et_2O) gave a solution that was washed with brine. The aqueous phase was extracted with Et₂O (30 mL), and the combined organic extracts were dried, filtered, and concentrated. Flash chromatography of the residue (6% EtOAc/ hexanes) afforded 96 mg of methyl ester 8 (65%, colorless oil) and 23 mg of the protonated product 10b (16%). Data for 8: ¹H NMR δ 5.57 (1 H, dd, $J \sim$ 6.9, 3.5 Hz, H-9), 3.67 (3 H, s, OCH₃), 2.45 (1 H, m, H-14), 2.28 (4 H, m, two H-11 and two H-24), 0.95 $(3 \text{ H}, d, J \sim 6.4 \text{ Hz}, C_{21}\text{-}CH_3), 0.76 (3 \text{ H}, \text{s}, C_{18}\text{-}CH_3); {}^{13}\text{C} \text{ NMR}$ δ 173.7 (CO₂), 149.7 (C-8), 121.0 (CF₃), 115.8 (C-9), 55.9, 53.8, 49.9, 44.9 (C), 35.4, 34.9 (CH₂), 34.6 (CH₂), 33.4 (CH₂), 27.9 (CH₂), 23.4 (CH₂), 21.1 (CH₂), 18.1, 10.8. Anal. Calcd for C₁₈H₂₇O₅F₃S: C, 52.42; H, 6.60. Found: C, 52.69; H, 6.27.

Des-*A*,*B***·25-oxo-27-norcholest-8-enyl trifluoromethane-sulfonate (19):** ¹H NMR δ 5.56 (1 H, dd, $J \sim 6.9$, 3.5 Hz, H-9), 2.40 (1 H, m, H-14), 2.30 (2 H, m, H-11), 2.14 (3 H, s, C₂₈-CH₃), 2.00 (2 H, m, H-24), 0.95 (3 H, d, $J \sim 6.4$ Hz, C₂₁-CH₃), 0.75 (3 H, s, C₁₈-CH₃); ¹³C NMR δ 209.1 (CO), 149.9 (C-8), 121.1 (CF₃), 116.0 (C-9), 54.0, 50.1, 45.2 (C), 44.0 (CH₂), 37.8, 35.2 (CH₂), 34.8 (CH₂), 29.8, 28.2 (CH₂), 23.7 (CH₂), 21.4 (CH₂), 20.2 (CH₂), 18.4, 11.2. Anal. Calcd for C₁₈H₂₇O₄F₃S: C, 54.53; H, 6.86. Found: C, 54.87; H, 6.83.

Des-*A*,*B*-8-[(trifluoromethanesulfonyl)oxy]-chol-8-ene-24-carbonitrile (20): ¹H NMR δ 5.58 (1 H, dd, $J \sim 6.9$, 3.5 Hz, H-9), 2.47 (1 H, m, H-14), 2.29–2.33 (4 H, m, two H-11 and two H-24), 0.96 (3 H, d, $J \sim 6.5$ Hz, 3 H, C₂₁-CH₃), 0.77 (3 H, s, C₁₈-CH₃); ¹³C NMR δ 149.7 (C-8), 120.0 (CF₃), 119.7 (CN), 116.1 (C-9), 53.9, 50.0, 45.1, 35.3, 34.7 (CH₂), 34.7 (CH₂), 28.1 (CH₂), 23.6 (CH₂), 22.0 (CH₂), 21.3 (CH₂), 18.2, 17.2 (CH₂), 11.1. Anal. Calcd for C₁₇H₂₄O₃F₃SN: C, 53.81; H, 6.37; N, 3.69. Found: C, 53.47; H, 6.20; N, 3.51.

Des-*A*,*B***-8-[(trilfuoromethanesulfonyl)oxy]chol-8-ene-**24-carboxamide (21): ¹H NMR δ 5.70 (br s, NH₂), 5.58 (1 H, dd, $J \sim 6.9, 3.5$ Hz, H-9), 2.50 (1 H, m, H-14), 2.29 (2 H, m, H-11), 2.18 (2 H, m, H-24), 0.96 (3 H, d, $J \sim 12.3$ Hz, C₂₁-CH₃), 0.76 (3 H, s, C₁₈-CH₃); ¹³C NMR δ 175.6 (CONH₂), 149.9 (C-8), 124.2 (CF₃), 116.1 (C-9), 54.0, 50.1, 45.2, 36.1, 35.7, 35.2, 34.8, 28.2, 23.8, 21.9, 21.6, 18.5, 11.2.

Des-*A*,*B***-24-(methanesulfonyl)chol-8-enyl trifluoromethanesulfonate (22):** ¹H NMR δ 5.58 (1 H, dd, $J \sim 6.9$, 3.5 Hz, H-9), 2.98 (2 H, m, H-24), 2.90 (3 H, s, SO₂CH₃), 2.46 (1 H, m, H-14), 2.30 (2 H, m, H-11), 0.98 (3 H, d, $J \sim 6.5$ Hz, C₂₁-CH₃), 0.76 (3 H, s, C₁₈-CH₃); ¹³C NMR δ 149.6 (C-8), 121.1 (CF₃), 116.1 (C-9), 55.0 (CH₂), 53.8, 50.0, 45.2 (C), 40.5, 35.6, 34.7 (CH₂), 34.4 (CH₂), 28.2 (CH₂), 23.7 (CH₂), 21.3 (CH₂), 19.0 (CH₂), 18.3, 11.2. Anal. Calcd for C₁₇H₂₇O₅F₃S₂: C, 47.21 H, 6.29. Found: C, 47.02; H, 6.13.

Des-*A*,*B***-24-(phenylsulfinyl)chol-8-enyl trifluoromethanesulfonate (23):** ¹H NMR δ 7.62 (2 H, m, o-H-Ph), 7.52 (3 H, m, *m*-2H-Ph and *p*-H-Ph), 5.57 (1 H, dd, $J \sim 6.9$, 3.5 Hz, H-9), 2.76 (2 H, m, H-24), 2.45 (1 H, m, H-14), 2.30 (2 H, *m*, H-11), 0.93 (3 H, d, $J \sim 6.5$ Hz, C₂₁-CH₃), 0.75 (3 H, s, C₁₈-CH₃); ¹³C NMR δ 149.8 (C-8), 144.3 (PhSO), 131.0 (PhSO), 129.3 (PhSO), 124.0 (PhSO), 121.1 (CF₃), 116.1 (C-9), 57.8, 54.0, 50.1, 45.2, 35.8, 34.8, 29.6, 28.2, 23.8, 21.4, 19.1, 18.9, 18.4, 11.3.

Procedure for Palladium-Catalyzed Coupling between Vinyl Triflates and Enynes: 1α -[(tert-Butyldimethylsilyl)oxy]-6,7-didehydro-24-(methoxycarbonyl)-25,26,27-trisnorprevitamin D₃ tert-Butyldimethylsilyl Ether (11). A mixture of enyne 5 (507 mg, 1.33 mmol), triflate 8 (500 mg, 1.213 mmol), Et₃N (0.67 mL, 4.85 mmol), and (Ph₃P)₂PdCl₂ (16 mg, 0.024 mmol, 2%) in DMF (21 mL) was heated at 75 °C for 1 h. The mixture was cooled to room temperature, diluted with EtOAc/hexanes (50 mL, 1:3), and washed with brine (70 mL). Drying, filtration, and concentration gave a dark yellow residue which was purified by flash chromatography (2-4% Et₂O/ hexanes) to afford 675 mg of dienyne 11 (86%, viscous liquid decomposing rapidly even at -10 °C but stable in hexanes in the refrigerator): ¹H NMR δ 5.97 (1 H, m, J ~ 3.5 Hz, H-9), 4.20 (1 H, m, H-1), 4.13 (1 H, m, H-3), 3.68 (3 H, s, OCH₃), 1.90 (3 H, s, C_{19} -CH₃), 0.96 (3 H, d, $J \sim 6.5$ Hz, C_{21} -CH₃), 0.89 (18 H, s, two t-BuSi), 0.70 (3 H, s, C18-CH3), 0.10 (6 H, s, Si(CH3)2), 0.07 (6 H, s, Si(CH₃)₂); ¹³C NMR δ 174.0 (CO₂), 140.2 (C), 133.4 (C-9), 122.6 (C), 115.5 (C), 92.4 (C), 88.1 (C), 69.9 (CH), 64.1 (CH), 54.4, 51.2, 50.0, 41.7 (C), 41.2 (CH₂), 39.7 (CH₂), 35.8, 35.2 (CH₂), 34.3 (CH₂), 27.8 (CH₂), 25.7 (CH₃), 25.7 (CH₃), 25.6 (CH₂), 25.0 (CH₂), 24.0 (CH₂), 21.4, 19.0, 18.5, 17.9, 17.8, 10.9, -4.5 (CH₃), -4.8 (CH₃), -4.9 (CH₃), -5.0 (CH₃).

Procedure for Hydrogenation with Lindlar Catalyst: 1a-[(tert-Butyldimethylsilyl)oxy]-24-(methoxycarbonyl)-25,26,27-trisnorprevitamin D₃ tert-Butyldimethylsilyl Ether (12). A solution of quinoline in hexanes (0.2 mL of a solution of 50 μ L of quinoline in 10 mL of hexanes) was added to a solution of dienyne 11 (305 mg, 0.475 mmol) in hexanes (12 mL). Lindlar catalyst (50 mg, previously high vacuum dried at 60 °C for 4 h) was added and the resulting solution was exposed to hydrogen gas at balloon pressure. After the solution was stirred for 8 h, TLC (2% EtOAc/hexanes) indicated that all the starting material had been converted to a product with a slightly higher R_f (monitoring by TLC is recommended in order to avoid overhydrogenation). After filtration and concentration, the residue was purified by flash chromatography (1-3% Et₂O/hexanes) to give 295 mg of protected previtamin D 12 (97%, colorless oil). This compound slowly equilibrates to the corresponding vitamin D at rt. In the ¹H NMR spectrum a small amount of the corresponding vitamin D form can be appreciated: ¹H NMR δ 5.87 and 5.72 (2 H, AB pattern, d, $J \sim 12$ Hz, H-6 and H-7), 5.52 (1 H, br s, J ~ 3.5 Hz, H-9), 4.15 (2 H, m, H-1 and H-3), 3.66 (3 H, s, OCH₃), $0.94 (3 H, d, J \sim 6.5 Hz, C_{21}$ -CH₃), 0.88 (9 H, s, t-BuSi), 0.86 (9 H)H, s, t-BuSi), 0.68 (3 H, s, C₁₈-CH₃), 0.08 (6 H, s, Si(CH₃)₂), 0.03 (6 H, s, Si(CH₃)₂); ¹³C NMR δ 174.7 (CO₂), 136.9 (C), 131.2 (C), 130.5 (C), 130.4 (CH), 129.2 (CH), 125.7 (C-19), 71.9 (CH), 65.6 (CH), 54.6, 51.8, 51.4, 42.7, 42.5, 39.7, 36.8, 36.5, 36.0, 35.0, 28.8, 26.3, 26.2, 25.5, 24.1, 22.2, 19.0, 18.6, 18.5, 17.9, 11.6, -4.0, -4.3, -4.4. -4.5.

Procedure for the Isomerization of Previtamins D to Vitamins D: 1a-[(tert-Butyldimethylsilyl)oxy]-24-(methoxycarbonyl)-25,26,27-trisnorvitamin D₃ tert-Butyldimethylsilyl Ether (13). Previtamin D 12 (295 mg, 0.458 mmol) was dissolved in isooctane (15 mL, distilled from Na) and refluxed in the dark for 5 h. Concentration gave a residue which was purified by flash chromatography (2-4% Et₂O/hexanes) to afford 290 mg of 13 (96%). A small amount of the previtamin D form in equilibrium is observed in the ¹H NMR spectrum: ¹H NMR δ 6.24 and 6.02 (2 H, AB pattern, d, $J \sim 11.2$ Hz, H-6 and H-7), 5.18 (1 H, m, $J \sim 3.5$ Hz, H-19E), 4.87 (1 H, m, $J \sim 3.5$ Hz, H-19Z), 4.37 (1 H, m, H-1), 4.18 (1 H, m, H-3), 3.67 (3 H, s, OCH_3 , 0.93 (3 H, d, $J \sim 6.2$ Hz, C_{21} -CH₃), 0.88 (18 H, s, two t-BuSi), 0.53 (3 H, s, C₁₈-CH₃), 0.07 (12 H, s, two Si(CH₃)₂; ¹³C NMR δ 173.1 (CO₂), 148.4 (C), 141.0 (C), 135.0 (C), 123.2 (CH), 118.0 (CH), 111.2 (C-19), 72.1 (CH), 67.5 (CH), 56.3, 51.3, 46.0, 45.7, 44.8, 40.6, 35.8, 35.3, 34.4, 31.5, 28.8, 27.6, 25.8, 25.7, 23.4, 22.6, 22.1, 21.5, 18.7, 18.1, 18.0, 14.0, 11.9, -4.8, -4.8, -4.9, -5.2. Anal. Calcd for C₃₈H₆₆O₄Si₂: C, 70.75; H, 10.62. Found: C, 70.42; H. 10.43

 1α ,25-Dihydroxyvitamin D₈ (1c). A solution of IMgCH₃ (0.33 mL, 1 M, 0.33 mmol) in Et₂O was slowly added to a cooled (-78 °C) solution of 13 (71 mg, 0.11 mmol) in THF (5 mL). The reaction mixture was warmed to rt and stirred for 10 h. The mixture was cooled to 0 °C, quenched with MeOH (several drops), poured into saturated aqueous NH₄Cl (15 mL), and extracted with Et₂O (50 mL). The organic extract was washed with brine (20 mL), dried, filtered, and concentrated. Filtration through a flash chromatography column (40% Et₂O/hexanes) gave a residue (62 mg) that was dissolved in THF (7 mL) and stirred at rt in the dark with TBAF (0.4 mL, 1 M in THF, 0.4 mmol) for 24 h. Concentration gave a solid residue that was diluted with EtOAc (15 mL), washed with brine (10 mL), dried, filtered, and concentrated. Flash chromatography of the residue (60% EtOAc/ hexanes) gave the hormone 1c (85 mg, 73% over the two steps) that was identical (NMR, TLC) to an authentic sample.^{14c}

26,27-Hexadeuterio-1 α ,25-dihydroxyvitamin D₃ (14).²⁰ See the procedure described for the preparation of 1c: ¹H NMR (CD₂Cl₂) δ 6.33 and 5.98 (2 H, AB pattern, d, $J \sim 12.4$ Hz, H-6 and H-7), 5.26 (1 H, m, H-19*E*), 4.93 (1 H, br s, H-19*Z*), 4.34 (1 H, m, H-1), 4.13 (1 H, m, H-3), 0.90 (3 H, d, $J \sim 6.2$ Hz, C₂₁-CH₃), 0.51 (3 H, s, C₁₈-CH₃); ¹³C NMR 148.7 (C), 143.5 (C), 133.9 (C), 125.1 (CH), 117.6 (CH), 111.8 (C-19), 71.2 (CH), 70.8, 67.2 (CH), 57.1, 56.8, 46.3, 45.8, 44.8, 43.5, 40.9, 36.9, 36.5, 29.4, 28.0, 24.0, 22.6, 21.2, 19.0, 12.1.

26.27-Dinor-25.25-dicyclopropyl-1a,25-dihydroxyvitamin D_3 (15). Cyclopropyl bromide (40 μ L, 0.51 mmol) was slowly added to a cooled (-20 °C) solution of t-BuLi in Et₂O (0.602 mL, 1.7 M. 1.02 mmol). The resulting mixture was warmed to rt (a white solid appeared) and diluted with Et₂O (2.4 mL). This solution (1 mL) was slowly added to a cooled (-78 °C) solution of 13 (50 mg, 0.077 mmol) in Et_2O (3 mL). The reaction mixture was allowed to come to -40 °C (4 h) and quenched with a few drops of water. The resulting solution was diluted with Et₂O, washed with brine (25 mL), dried, filtered, and concentrated. The concentrate was filtered through a flash chromatography column (2% Et₂O/hexanes), affording a product (46 mg) which was dissolved in THF (7 mL) and stirred in the dark at rt with TBAF (0.36 mL, 1 M in THF, 0.36 mmol) for 24 h. Concentration gave a residue which was diluted with EtOAc (20 mL). The solution was washed with brine (10 mL), dried, filtered, concentrated, and flash chromatographed (60 $\%\,$ EtOAc/hexanes) to give 23 mg of the desired compound 15 (65% over the two steps, white solid, mp 59 °C): ¹H NMR (CD₂Cl₂) δ 6.44 and 5.99 (2 H, AB pattern, d, J ~ 11 Hz, H-6 and H-7), 5.27 (1 H, brd, J ~ 3.5 Hz, H-19E), 4.95 (1 H, brd, $J \sim 3.5$ Hz, H-19Z), 4.35 (1 H, m, H-1), 4.15 (1 H, m, H-3), 0.92 (3 H, d, J ~ 6.2 Hz, C₂₁-CH₃), 0.81 (2 H, m, H-26 and H-27), 0.53 (3 H, s, C₁₈-CH₃), 0.34 (8 H, m, 4-cyclopropylic CH₂); ¹³C NMR (CD₂Cl₂) δ 148.6 (c), 143.5 (c), 133.9 (C), 125.1 (CH), 117.6 (CH), 111.8 (C-19), 71.2 (CH), 71.0 (CH), 67.2, 57.2, 56.8, 45.8 (C-25), 43.5 (CH₂), 43.4 (CH₂), 41.0 (CH₂), 37.1 (CH₂), 36.6, 29.4 (CH₂), 28.0 (CH₂), 24.0 (CH₂), 22.6 (CH₂), 20.8 (CH₂), 19.0 (CH), 12.1, 0.8 (CH₂), -0.5 (CH₂).

26,27-Dinor-25,25-di-tert-butyl-1a,25-dihydroxyvitamin D₃ (16). A solution of t-BuLi in Et₂O (0.123 mL, 1.7 M, 0.21 mmol) was added to a cooled (-78 °C) solution of 13 (45 mg, 0.97 mmol) in $Et_2O(5 mL)$. During the addition the reaction mixture changed from colorless to brown. The mixture was allowed to come to rt, and after stirring for 1.5 h was cooled to 0 °C, quenched by addition of a few drops of water, diluted with Et₂O (50 mL), washed with brine (20 mL), dried, filtered, and concentrated. The resulting white solid was dissolved in THF (8 mL). The solution was stirred in the dark at rt with TBAF (0.56 mL, 1 M in THF, 0.56 mmol) for 16 h. Concentration gave a residue which was diluted with EtOAc (20 mL), washed with brine (10 mL), dried, filtered, concentrated, and flash chromatographed (60% EtOAc/hexanes) to give 19 mg of 16 (68% over the two steps, colorless viscous oil): ¹H NMR δ 6.38, and 6.01 (2 H, AB pattern, d, $J \sim 11$ Hz, H-6 and H-7), 5.33 (1 H, m, H-19E), 5.00 (1 H, m, H19Z), 4.43 (1 H, m, H-1), 4.23 (1 H, m, H-3), 1.00 (18 H, s, two t-Bu), 0.93 (3 H, d, $J \sim 6.2$ Hz, C₂₁-CH₃), 0.54 (3 H, s, C₁₈-CH₃); ¹³C NMR δ 148.7 (C), 143.4 (C), 134.0 (C), 125.0 (CH), 117.6 (CH), 111.8 (C-19), 80.0 (C-25), 71.1 (CH), 67.1 (CH), 60.6, 56.9, 56.8, 46.3 (CH₂), 45.8, 43.4 (CH2), 42.8, 40.9 (CH2), 37.0 (CH2), 36.5, 34.2 (CH2), 29.4 (CH₂), 28.8, 28.0 (CH₂), 24.0 (CH₂), 23.3 (CH₂), 22.7 (CH₂), 19.2. 12.1.

26,27-Dinor-25,25-diphenyl-1 α ,**25-dihydroxyvitamin D₃ (17).** Bromobenzene (53.3 μ L, 0.51 mmol) was slowly added to a cooled (-70 °C) solution of *t*-BuLi in Et₂O (659 μ L, 1.49 M). The reaction mixture was warmed to rt and diluted with Et₂O (1.3 mL), and the solution (1 mL) was added to a cooled (-78 °C) solution of 13 (70 mg, 0.11 mmol) in Et_2O (4 mL). The reaction mixture was stirred for 30 min, quenched with several drops of water, diluted with EtOAc/hexanes (20 mL 1:3), washed with saturated aqueous NH4Cl (15 mL) and brine (25 mL), dried, filtered, and concentrated. The residue was filtered through a flash chromatography column (4% Et₂O/hexanes). The product (80 mg) was dissolved in THF (7 mL) and stirred in the dark at rt with TBAF (0.67 mL, 1.1 M in THF, 0.85 mmol) for 20 h. Concentration gave a residue which was diluted with EtOAc (20 mL), washed with brine (10 mL), dried, filtered, concentrated, and flash chromatographed (60% EtOAc/hexanes) to give 43 mg of 17 (71% over the two)steps, mp 128 °C); ¹H NMR (CD₂Cl₂) δ 7.40 (4 H, d, J ~ 8 Hz, o-4H-Ph), 7.31 (4 H, t, J ~ 7 Hz, m-4H-Ph), 7.18 (2 H, t, J ~ 7 Hz, p-H-Ph), 6.34 and 5.99 (2 H, AB pattern, J ~ 11.5 Hz, H-6 and H-7), 5.28 (1 H, br s, H-19E), 4.94 (1 H, br s, H-19Z), 4.35 (1 H, m, H-1), 4.15 (1 H, m, H-3), 0.83 (3 H, d, $J \sim 6.5$ Hz, C₂₁-CH₃), 0.50 (3 H, s, C₁₈-CH₃); ¹³C NMR (CD₂Cl₂) δ 148.6 (C), 148.0 (C), 143.4 (C), 133.9 (C), 128.5 (CH), 127.1 (CH), 126.4 (CH), 125.0 (CH), 117.6 (CH), 111.8 (C-19), 78.5 (C), 71.2 (CH), 67.2 (CH), 57.1, 56.7, 46.2, 45.7 (CH2), 43.4 (CH2), 42.7 (CH2), 40.9 (CH₂), 36.6 (CH₂), 36.4, 29.4 (CH₂), 28.0 (CH₂), 24.0 (CH₂), 22.6 (CH₂), 20.7 (CH₂), 18.9, 12.1.

 1α -[(*tert*-Butyldimethylsilyl)oxy]-6,7-didehydro-24-(methanesulfonyl)-25,26,27-trisnorprevitamin D₃ *tert*-Butyldimethylsilyl Ether (24). This compound was prepared from triflate 22 and enyne 5 following the same procedure used for the preparation of 11: ¹H NMR δ 5.97 (1 H, m, H-9), 4.19 (1 H, m, 1H, H-1), 4.10 (1 H, m, H-3), 2.95 (2 H, m, H-24), 2.91 (3 H, s, SO₂CH₃), 0.99 (3 H, d, J ~ 6.5 Hz, C₂₁-CH₃), 0.89 (9 H, s, *t*-BuSi), 0.88 (9 H, s, *t*-BuSi), 0.71 (3 H, s, C₁₈-CH₃), 0.10 (6 H, s, Si(CH₃)₂), 0.07 (6 H, s, Si(CH₃)₂); ¹³C NMR δ 140.5 (C), 133.2 (CH), 122.5 (C), 115.5 (C), 92.3 (C), 82.3 (C), 70.0 (CH), 64.2 (CH), 52.3, 54.3, 50.1, 41.9, 41.2, 40.5, 39.8, 35.8, 34.6, 27.9, 25.8 (CH₃), 25.7 (CH₃), 25.0, 24.1, 19.1, 19.1, 18.4, 18.0, 18.0, 11.0, -4.4 (CH₃), -4.7 (CH₃), -4.8 (CH₃), -4.9 (CH₃).

24-(Methanesulfonyl)-1 α ,25-dihydroxy-25,26,27-trisnorvitamin D₃ (18). Partial hydrogenation and thermal isomerization were carried out following the same procedures used for the preparation of 13 from 11. Desilylation was carried out as above. Compound 18 (colorless oil) was obtained in a 73% yield over the three steps. ¹H NMR δ 6.34 and 5.98 (2 H, AB pattern, $J \sim 11.2$ Hz, H-7 and H-6), 5.29 (1 H, br s, H-19*E*), 4.94 (1 H, br s, H-19*Z*), 4.35 (1 H, m, H-1), 4.15 (1 H, m, H-3), 2.88–2.97 (2 H, m, H-24), 2.84 (3 H, s, SO₂CH₃), 0.94 (3 H, d, $J \sim 6.4$ Hz, C₂₁-CH₃), 0.53 (3 H, s, C₁₈-CH₃); ¹³C NMR δ 148.6 (C), 143.2 (C), 134.1 (C), 125.0 (CH), 117.0 (CH), 111.8 (CH19), 71.2 (CH), 67.2 (CH), 56.7, 56.6, 55.7, 53.5, 46.3, 45.8, 43.4, 40.9, 36.2, 35.0, 29.4, 28.0, 23.9, 22.6, 19.7, 18.8, 12.1.

Acknowledgment. Support for this research under Grant PB90-0759 from the DGICYT (M.E.C., Spain) is gratefully acknowledged. J.P.S. thanks the Xunta de Galicia for a fellowship. We also thank Duphar for the generous gift of vitamin D_2 used for the preparation of Lythgoe-Inhoffen diol (6a).

Supplementary Material Available: Spectral and analytical data (¹H NMR, ¹³C NMR, IR, UV, MS, HRMS, and other analytical data) and ¹H and ¹³C NMR spectra (19 pages). This material is contained in libraries on microfiche, immediately follows this article in the microfilm version of the journal, and can be ordered from the ACS; see any current masthead page for ordering information.